

VOLUME 8 ISSUE 1 2022

ISSN 2454-3055



**INTERNATIONAL
JOURNAL OF
ZOOLOGICAL
INVESTIGATIONS**

***Forum for Biological and
Environmental Sciences***

Published by Saran Publications, India



International Journal of Zoological Investigations

Contents available at Journals Home Page: www.ijzi.net

Editor-in-Chief: Prof. Ajai Kumar Srivastav

Published by: Saran Publications, Gorakhpur, India



ISSN: 2454-3055

Efficacy of Phytochemicals Against Mosquito Larvae: An Update to Integrated Mosquito Management

Kumar Sanjeev and Kumar Sushil*

Department of Zoology, Deen Dayal Upadhyaya Gorakhpur University, Gorakhpur 273009, India

*Corresponding Author

Received: 28th January, 2022; Accepted: 10th March, 2022; Published online: 15th March, 2022

<https://doi.org/10.33745/ijzi.2022.v08i01.034>

Abstract: Mosquitoes are considered a most common human enemy; causes mortality, morbidity, monetary and social disruption. It plays a significant role in endemic conditions for Malaria, Japanese encephalitis (JE), Dengue, Yellow fever, Chikungunya, and Filariasis etc. The prolonged use of synthetic insecticides raises the risk of unintended harmful or fatal consequences such as biological amplification, mosquito resistance development, and negative impacts on environmental quality and non-target species, including human health. So that there is urgent need to find natural and environmentally safe phytochemicals as an alternative to synthetic pesticides from indigenous plant sources that are accessible to the whole local community. Phytochemicals are botanicals that are naturally occurring pesticides produced from floral resources, and they are a viable alternative to synthetic insecticides in Integrated Mosquito Management (IMM). Plant based insecticides are less toxic, delay the development of resistance because of its structure and easily biodegradable. In this article, we have summarized the recent studies published in 2020 and 2021 on the biopesticide effect of botanicals with their suitable solvents which possess strong potential against mosquitoes larva. We also focused on their mode of action on the target population, variance in larvicidal efficacy among *Culex*, *Aedes*, and *Anopheles* mosquitoes, polarity of solvents employed during extraction, active component, and prospective improvements in biological control.

Keywords: Synthetic insecticides, Phytochemicals, Larvicidal, Secondary metabolites, LC₅₀, *Culex quinquefasciatus*, *Aedes aegypti*, *Anopheles stephensi*

Citation: Kumar Sanjeev and Kumar Sushil: Efficacy of phytochemicals against mosquito larvae: An update to integrated mosquito management. Intern. J. Zool. Invest. 8(1): 305-319, 2022.

<https://doi.org/10.33745/ijzi.2022.v08i01.034>

Introduction

Mosquitoes are considered as oldest human enemies and, play a significant role in the health hazards to society. They transmit many medically important pathogens and parasites such as bacteria, viruses, protozoans and roundworms (Sathishkumar and Manimegalai, 2008). They have the ideal ability to transfer the vector pathogens that affect millions of people

worldwide, especially in developing countries due to their unhygienic and poorest conditions (WHO, 1992; James, 1992; Awad and Shimaila, 2003). World Health Organization (WHO) has declared that mosquito (Diptera: Culicidae) is a "Public enemy number one" as they act as a vector for fatal diseases like Malaria, Dengue, Chikungunya, Japanese Encephalitis (JE), Filariasis, Yellow fever,

Lyme disease, Poly arthritis etc. globally including India (WHO, 1996; Gubler *et al.*, 1998; Ghosh *et al.*, 2012; Kalita *et al.*, 2013; Dawet *et al.*, 2016). If we see the epidemiology of these diseases, it's a big source of malaria with an estimated 241 million new cases and 627,000 deaths worldwide in 2020 due to unfocused prevention strategies of malaria during the pandemic of COVID-19 (WHO, 2021). Another mosquito borne infection, dengue, covers 390 million people including 70% of the actual burden in Asia, including India, which has reported 32,796 cases upto November 30, 2020 (WHO, 2020). There are about 68,000 clinical cases of encephalitis, 13,600-20,400 deaths per year globally, and more than 3 billion people at risk of infection (WHO, 2020). JE cases have recently declined; it showed 44,000 new cases and 6,000 fatalities between 2008 to 2014, particularly in the states of Uttar Pradesh and Bihar, India. In 2016, encephalitis cases rise with over 125 children reported to have died in one hospital in Gorakhpur (Times of India, 2016). As of November 30, 2020, India has reported 32,287 suspected cases and 5,159 confirmed cases of chikungunya (WHO, 2020). There are also about 893 million people in 49 countries infected by filariasis, including 23 states known to be endemic to lymphatic filariasis in India (WHO, 2020).

Integrated mosquito management (IMM) is based on knowledge of mosquito biology, their life cycle, and vector phenomenon of mosquitos. There are about 3500 species of mosquitoes, classified into three subfamilies, named as Toxorhynchitinae, Anophelinae, and Culicinae (Reinert, 2000; Bayer Environmental Science, 2007). *Culex*, *Aedes*, and *Anopheles* genera constitute the major mosquito vectors, in which mostly hazardous mosquitoes are *Culex quinquefasciatus*, *Aedes aegypti*, and *Anopheles stephensi* which transmitted many endemic diseases in India including this lowland area recognized as a Poorvanchal (Redwane *et al.*, 2002). This region provides a better breeding place for mosquitoes with the significant presence of poor drainage systems, an increasing rate of mosquitoes in fish ponds, especially in the rainy

season, irrigation ditches, and rice fields containing stagnant water. Thus, it is essential to control mosquitoes to prevent vector borne diseases and their transmission. The approaches to disrupting disease transmission by above vectors are the killing of mosquitoes and preventing mosquito's bites (Malik *et al.*, 2013; Kumar *et al.*, 2012). Synthetic pesticides are used for public health sprays as chemicals with insecticidal properties such as organochlorines, organophosphorus, carbamates, pyrethroids, synthetic pyrethroid pyrroles, and phenyl pyrazole (Schofield, 1993; Pal, 1994). The excessive, unlimited, uninterrupted, indiscriminate, and continuous application of synthetic insecticides as principal agents results in the unwarranted toxic or lethal effects on non-target organisms, the development of resistance in mosquitoes, and more importantly, potential toxicity in the environment and adverse effects on human health, posing a great threat to life and the environment (Shrivastava *et al.*, 2002; Kawada *et al.*, 2009; Koures *et al.*, 2012; Raveen *et al.*, 2014; Rathy *et al.*, 2014; Hemlatha *et al.*, 2015; Roy *et al.*, 2017). So, there is an urgent need for an alternative source to synthetic insecticides across the globe.

Phytochemicals from natural resources are emerging as a promising alternative to synthetic insecticides for insect management. They are effective, environment friendly, easily biodegradable and have less or no effect on non-target organisms. They are also often less expensive than synthetic insecticides (Govindarajan *et al.*, 2011). Different chemical classes of phytochemicals are investigated for insect control potential and found promising, such as steroids, alkaloids, terpenes, and phenolic constituents (Nisha *et al.*, 2009; Pavela 2015; Pavela and Benelli 2016; Pavela *et al.*, 2017). Essential oils are complex mixtures of phytochemicals extracted from citronella plants (citronellal), neem (azadirachtin), lavender (linalool), and mentha (p-menthane) plants (Chaubey, 2019). Their biological effects might be the result of a synergistic effect involving all the

molecules, or they could merely reflect the actions of the major molecules. However, the ability of phytochemicals to repel insects varies with age, species, plant source of extract, collection site and solvent used for extraction (Isman, 2015; Benelli 2015). Many investigations have shown the efficacy of different solvent extracts of plants and essential oils as mosquito larvicides, as repellents, ovideterrents and growth inhibitor without causing any harm to humans (Pavela 2015; Govindarajan *et al.*, 2016a, b, c, d, e; da Silva *et al.*, 2016; Pavela and Benelli, 2016; Benelli *et al.*, 2018; Lalthazuali and Nisha, 2017). These bioactive chemicals may act as insecticides, repellents, antifeedants, moulting hormones, juvenile hormones mimics, growth inhibitors, anti-moulting hormones, as well as attractants. Plant based insecticides are less toxic, delay the development of resistance because their structures are easily biodegradable (Ignacimuthu, 2000). Approximately 2000 species of terrestrial plants are reported with their insecticidal properties including plant's secondary metabolites and their derivatives which provides substitute source for reducing the mosquito population (Feinstein, 1952; Shivkumar *et al.*, 2013; Kumar *et al.*, 2021). Recently, investigations have proved the effectiveness of phytochemical compounds such as saponine, steroids, isoflavonoids, essential oils, alkaloids, and tannins against mosquito larvae (Cavalcanti *et al.*, 2004; Wiseman and Chapagain, 2005; Khanna and Konnabrian 2007; Chowdhury *et al.*, 2008; Joseph *et al.*, 2008). It can be easily eradicated in their larval stages for mosquito control operations in comparison to adults.

We have summarized the major studies on phytochemicals that have been effective on larvae, adults or both types of mosquitoes. This review is focused on the basic methods of mosquito vector control, synthetic insecticides and their drawbacks, the role of phytochemicals with their sources and modes of action and the current situation of natural plant sources for reducing the risk of mosquito borne diseases under the umbrella of Integrated Mosquito Management (IMM).

Integrated Mosquito Management (IMM) – An inclusive control approach:

IMM involves a combination of techniques and systems for keeping up with low levels of mosquito vectors. It is a dynamic cycle for the administration of mosquito populations. The motivation behind IMM is to provide insurance against sicknesses spread by mosquitoes, keep up with a healthy environment through legitimate use and removal of pesticides and work on general satisfaction through reasonable and successful mosquito control procedures. The major approaches of integrated vector control for mosquitoes are anti-larval, anti-adult, protection against mosquito bites, and legislative controls. Control of mosquitoes at an immature stage is necessary and efficient in IMM (Elimam *et al.*, 2009). Controlling mosquitoes at the larval stage is more efficient and target specific of all the avenues of IMM because the mosquitoes are relatively immobile during the immature stage; remaining the most concentrated than they are in the adult stage.

Mosquito larva control measures:

Mosquito vector control is the serious concern in developing countries like India due to the lack of general awareness, the development of resistance, and socio-economic reasons. Vector control, which includes both anti-larval and anti-adult measures, constitutes an important aspect of any mosquito control programme. In particular, larval habitats are small, widely dispersed and transient. So, larva control would seem to be an ideal approach to mosquito control as it eliminates mosquitoes before they reach the stage where they can transmit vector borne diseases. The major methods that may be implemented through source reduction, chemical larviciding, and biological control (Fig. 1) are described below:

Source Reduction:

The removal or permanent destruction of breeding grounds is referred to as "source reduction." Mosquito larvae habitats can be eradicated by filling water collecting depressions,

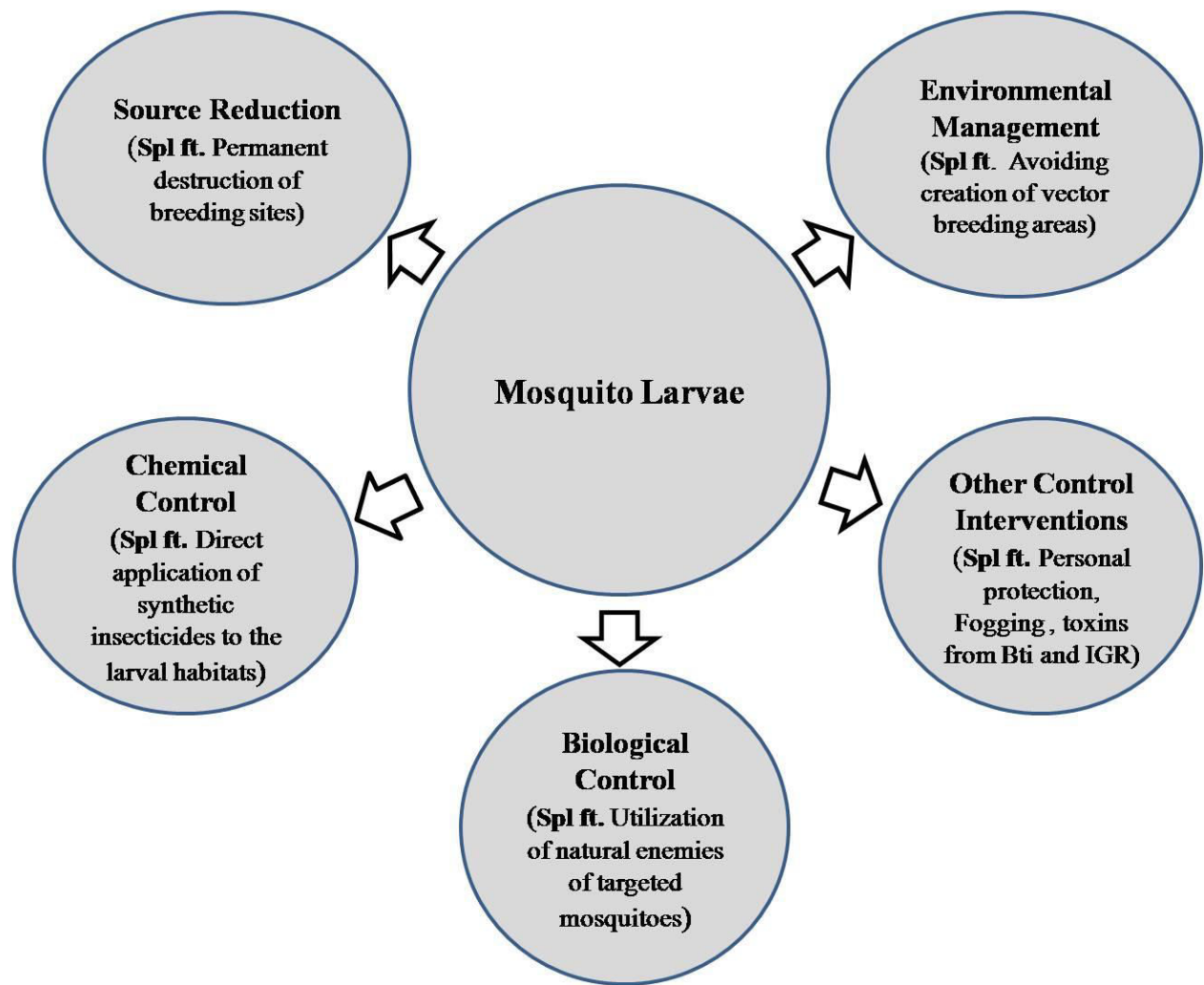


Fig. 1: Major methods of Mosquito's control. Spl ft.- special feature

swamps, or ditching marshy regions to remove stagnant water (Lawler and Lanzaro, 2005). Mosquitoes that breed in containers are more susceptible to sources because people can remove or cover stagnant water in cups, cans, and rain barrels around houses. Irrigation water-breeding mosquitoes can be reduced with careful water management.

Chemical application:

Some mosquito species are not controlled by habitat elimination alone but also need direct application of synthetic insecticides to the larval habitats. Chemical application methods include space spraying, larviciding, and targeted residual spraying. It can be planned fast, effectively, and generate outcomes at a cheap cost. Synthetic

insecticides have a unique function in mosquito-borne illness control programs, particularly in the initial stages of intervention when other control strategies are still developing and playing effective roles in an integrated strategy (Chavassee and Yap, 1997).

Biological Control:

To accomplish efficient vector management, biological control strategies involve the use of natural enemies of targeted mosquitoes as well as biological toxins. It includes alternative control measures such as invertebrate predators, nematodes (e.g., *Romanomermis culicivorax*), larvivorous fish (e.g., *Gambusia affinis*), including some protozoa and fungi (e.g., *Lagenidium giganteum*) (Das and Amalraj, 1997). They are

typically most feasible with easily identifiable breeding places.

Environmental management:

Environmental management is commonly used to lessen the burden of vector borne diseases over time. These include marsh alteration, house screening, filling, grading, and drainage of stagnant water places as well as vegetative plantings (Ault, 1994; Utzinger *et al.*, 2001). These applications aim to minimize the quantity of a target vector by avoiding the establishment of vector breeding places, modifying natural habitats, or improving human habitation while minimizing negative environmental and social impacts.

Other vector control interventions:

Apart from all the above measures, fogging also, called area spray, is used mostly in emergency situations such as epidemics (Agosta, 2002). Though, in any malaria endemic area, mosquitoes are typically found in regions where pesticides are difficult to reach and here fogging become not much more beneficial. Fogging must be timed to coincide with peak adult mosquito activity. It will need to be done regularly to have an impact and this might become too expensive to sustain or result in insecticide misuse. The use of oils added to the water surface can be utilized to suffocate the larvae and pupae of mosquitoes. Toxins from the bacterium *Bacillus thuringiensis* var. *Israelensi* and insect growth regulators like methoprene are mosquito-specific and can be used as effectively as chemical insecticides (Barnard, 1999). We must also include personal protection measures against mosquitoes such as window screens, insecticide-treated bed nets (ITNs) and repellents (such as N, N diethyl-meta-toluamide), including wearing habits of light coloured and full-sleeved clothes.

Other measures like ovitraps, space spray, biological control agents, etc. are examples of little development for mosquito control encouraging when used on a limited scale (Raghavendra *et al.*, 2011). Several biological agents recently introduced for vector control, such as insect growth regulators, biocontrol agents, and natural

plant products, have yet to gain the needed scale of utility, including bacterial pesticides (Hardin and Jackson, 2009). Environmental management has shown sufficient promise for vector and disease control, but still needs advocacy for intersectoral coordination and can sometimes work very intensive. Recently, genetic engineering and sterile insect technology have been developed and are being considered for use in routine vector control. Standardized procedures and methods are available, but with biological, ethical considerations, and an understanding of a properly trained workforce for implementation is required. They are technically focused methods. In particular, plants are extracted as an alternative form of synthetic chemicals for combating mosquito resistance problems.

Synthetic pesticide control measures: historical prospective and drawbacks:

Throughout history, scientists have experimented with numerous strategies to address the threat of mosquito-borne diseases with synthetic insecticides serving as the primary instruments in mosquito control operations. However, the widespread and repeated use of synthetic organic insecticides such as organochlorines and organophosphates have disrupted biological control systems. This may result in target species' revival and resistance as well as the destruction of non-target beneficial fauna, raising environmental and human health concerns. Pesticide residues have been shown to biomagnify when they enter the ecosystem and circulate through the food web. The negative consequences of insecticide use have required research and development of environmentally safe, biodegradable, and indigenous mosquito control methods.

The first incidence of DDT resistance was discovered in *Aedes tritaeniorhynchus* and *Aedes sollicitans* in 1947, not long after DDT was introduced for mosquito control in 1946 (Brown 1986). Since then, roughly half of more than 100 mosquito species have been reported as resistant to one or more insecticides (WHO, 1992). The principal categories of insecticides used for

malaria control are organochlorines, organophosphates, carbamates, and pyrethroids, with the latter increasingly taking a larger share of the market for both indoor residual spraying and large-scale insecticide-impregnated bednet programmes (Zaim and Guillet, 2002). Unfortunately, resistance tends to follow changes in insecticides. Key vectors of malaria like *Anopheles culicifacies*, *Anopheles stephensi*, *Anopheles albimanus*, *Anopheles arabiensis*, and *Anopheles sacharovi* have been reported to exhibit organophosphate (OP) resistance, which can be either broad-spectrum or malathion-specific OP (Eshgy 1977; Hemingway and Georgeon, 1983; Hemingway 1983, Hemingway *et al.*, 1985; Herath *et al.*, 1986). In addition, *Anopheles albimanus*, *Anopheles stephensi* and *Anopheles gambiae* have shown resistance to pyrethroids, while resistance to carbamates has been noted in *Anopheles sacharovi* and *Anopheles albimanus* (Brogdon and Barber, 1990; Hemingway *et al.*, 1992; Vatandoost *et al.*, 1996). All of the above mentioned studies and given the recent emphasis by the WHO, synthetic resistance has become the magnifier of mosquito-borne diseases worldwide, including developing countries like India. While synthetic pesticides are effective in combating mosquitoes, they cause many environmental problems, including pesticide resistance, pollution, and toxic side effects on humans. (Mecedo *et al.*, 1997; Lixi *et al.*, 2006). These problems, together with the insecticide residuals in the environment, pose us to seek attention towards alternative methods.

In past DDT sprays have had remarkable success in reducing disease-carrying organisms, but have led to the development of pesticide resistance. Organophosphates, carbamate, and synthetic pyrethroids were also introduced into indoor residue sprays with sufficient success to control resistance, but they subsequently developed extensive multi-insecticide resistance in vectors. As part of IMM, the search for novel bioactive compounds in plants was initiated after experiencing several issues through the injudicious and overuse of synthetic insecticides in

nature from the mid 1970s to till date. Efforts have been made to determine plant derived phytochemicals' structures and their commercial production for mosquito control.

Role of Phytochemicals in IMM:

Phytochemicals are botanicals that can be clearly based on pesticides acquired from floral resources. Many herbal products have been investigated and utilised as biopesticides, even before the discovery of synthetic pesticides botanicals such as *Pyrethrum*, *Chrysanthemum*, *Derris*, *Nicotine*, *Quassia*, *Azadirachtin*, and *Turpentine*, etc. (Shalaan *et al.*, 2005). Plants produce a variety of chemicals, many of which have medicinal and pesticide properties. More than 2,000 plant species are known to produce chemical factors and metabolites that are important for pest management programs. However, the discovery of synthetic insecticides, particularly organochlorines, such as DDT in 1939, diverted attention away from the use of phytochemicals in mosquito control (Shalaan *et al.*, 2005). After experiencing a number of issues due to imprudent and overuse of synthetic insecticides in nature, a refocus on phytochemicals that are readily biodegradable and have no adverse effects on non-target organisms was welcomed. Phytochemicals account for up to 1% of the global pesticide market (Isman, 1997).

Phytochemicals are largely secondary metabolites that help plants endure herbivorous predators' continual selection pressures and other environmental conditions. It has various plant sources, such as alkaloids, steroids, terpenoids, essential oils, and phenolics, which have previously been recommended for their insecticidal properties (Shalaan *et al.*, 2005). They have been extracted from the full plants or from various parts such as, roots, stems, bark, leaves, fruits, etc. In particular, members of the Solanaceae, Asteraceae, Zinniaceae, Laviatae, Milaceae, Oocystaceae, and Rutaceae families plants have different types of larvicidal, adulticidal or repellent activity against different mosquito

species (Shalaan *et al.*, 2005). A wide range of phytochemicals from herbs, shrubs, and large trees have been used to extract mosquito toxins.

Phytochemicals with mode and site of action for larvicidal activity:

Worldwide plants involve a rich, undiscovered pool of phytochemicals that might be utilized in place of synthetic insecticides in mosquito control. The mode and site of action of larvicidal phytochemicals, including essential oils, has received little attention. Kishore *et al.* (2011) surveyed the adequacy of phytochemicals against mosquito hatchlings as indicated by their substance nature and depicted the mosquito larvicidal probability of a few plant derived secondary materials, for example, alkanes, alkenes, alkynes, and basic aromatics, lactones, essential oils and unsaturated fats, terpenes, alkaloids, steroids, isoflavonoids, pterocarpanes, and lignans. They additionally archived the disengagement of a few bioactive toxic standards from different plants and revealed their harmfulness against various mosquito species (Kishore *et al.*, 2011). However, it is clear that they change the physiology of insects in a variety of ways by affecting biochemical processes in insects, especially disrupting the endocrine balance (Rattan, 2010). Monoterpenes, cineole, eugenol, limonene, terpinene, citronellol, citronellal, camphor, and thymol are incessant fixings in various essential oils that have mosquito repellent properties (Nerio *et al.*, 2010). The repellent phytochemicals connect with the female mosquito's olfactory receptors, hindering the impression of smell thus keeping the mosquitoes from perceiving the host (Tripathi *et al.*, 2009). It can also disrupt correspondence in mating conduct of bugs by impeding the capacity of antennal sensilla, and fruitless mating could prompt a sweetheart fertility and eventually bring down the number of inhabitants in the population of insect pests. However, botanicals primarily influence the midgut epithelium and optionally influence the gastric caeca and malpighian tubules of mosquito larvae (Rey *et al.*, 1999; David *et al.*,

2000). There has been report of hindering of acetyl cholinesterase (AChE) amalgamation by essential oils which assume part in cholinergic neural connections in insect and higher animals for nerve conduction (Fournier and Mutero, 1994). In the case of larvicidal activity, efficacy and insecticidal potency of phytochemicals seem to be dependent on relative absorption on the insect surface (Enan, 2001). However, it could be attributed to different species of mosquitoes or variation in composition of essential oils used (Lee *et al.*, 2002; Pavela, 2005). The mixture of phytochemicals also ought to be considered as an elective insect spray possibility for mosquito control in rearing locals, particularly in situations where helplessness is diminishing.

Larvicidal potential of phytochemicals depends on mosquito type, plant components, and solvent polarity:

Plant species, plant parts used, maturity of plant parts, solvent employed during extraction, and accessible mosquito vector species can all influence the effectiveness of phytochemicals against larvae. Changes in the larvicidal efficiency of plant extracts also happened as a result of a plant's geographical origin. It has been demonstrated that the polarity of the solvents employed affects the extraction of active biochemicals from plants, with polar solvents extracting polar compounds and non-polar solvents extracting non-polar molecules. Changing in solvent types can have a big impact on the potency of extracted phytochemicals, and plant species have varied chemo-profiles.

The best way to check the toxicity effect of phytochemicals is to use lethal concentration 50 (LC₅₀). It is the concentration of a compound that kills 50% of the tested population. In this study, we have covered updated studies, especially those published in the years 2020 and 2021 in the COVID-19 era. Out of 25 studies, only 10 studies met the qualifying inclusion criteria, which are mentioned here in context of target mosquitoes, plant name with family and used part for extraction, solvents and the level of LC₅₀ (Table 1).

Sogan *et al.* (2020) reported the lowest value of LC₅₀ as 6.843 recorded for leaf of *Spergula arvensis* (family: Caryophyllaceae) in the solvent of methanol against the *Anopheles culicifacies* (principal vector of Malaria). On other hand, Shahat *et al.* (2020) have recorded the highest LC₅₀ value of 683.28 in the same metholic solution of leaf extract of *Otostegia fruticosa* (family: Lamiaceae) aganisnt *Culex pipiens* (principal vector of West Nile virus). Different investigations additionally revealed the LC₅₀ level of various parts of plants among dissolvables utilised for extraction based on polarity: aqueous, methanol, ethanol, chloroform, hexane, ethyl acetic acid, and petroleum ether derivation against different types of mosquito's larvae (Table 1).

Isolation of toxic larvicidal active phytochemicals and a futuristic approach:

More land is required to support development, agriculture, or urbanization as the world's human population continues to expand at a rate of 1.24% each year (Melorose *et al.*, 2015). These human activities contribute to the opening up of possible mosquito breeding areas as well as the speciation of mosquito-borne disease vectors. Mosquito control has been fully reliant on synthetic organic compounds since the discovery of DDT. However, overuse of synthetic organic pesticides has resulted in environmental risks as well as the development of physiological resistance in important vector species during the last seven decades. Following repeated application, the overwhelming development of vector resistance to currently available synthetic pesticides continues to pose a significant threat to the transmission of diseases by vectors. This has forced the search again for the development of ecologically friendly, biodegradable, low-cost, indigenous vector control technologies that may be employed with little caution by individuals and communities in certain situations.

Although several plant derivatives have been found to exhibit mosquitocidal or repellant action against mosquito vectors, only a few plant

products have proven useful in mosquito control. Even before the discovery of synthetic organic insecticides, some phytochemicals and their derivatives, such as nicotine derived from tobacco leaves, *Nicotiana tabacum*, Anabesine and lupinine, the alkaloids extracted from Russian weed *Anabasis aphylla*, rotenone derived from *Derris eliptica*, and pyrethrums derived from *Chrysanthemum cinererifolium* flowers, were used as natural insecticides (Jacobson and Crosby, 1971; Campbell *et al.*, 1993). As one strategy, phytochemicals with proven pesticide or repellent capability can play a substantial role in interrupting the spread of mosquito-borne diseases at the personal and community level. The biologically active phytochemicals have yet to be tested on a significant scale for managing insect pests and vectors. As a result, a "laboratory to grassland" transfer of tested bioactive chemicals and subsequent incorporation into IMM might counteract pesticide resistance, significantly lowering vector populations and the danger of vector-borne disease transmission. It necessitates the availability of scalable and long-term technology for both small and large-scale manufacturers. Nanotechnology, in particular, is actively transforming the manufacture of commercial pesticides. It has revolutionized the manufacturing of plant-based silver nanoparticles, which have been shown to be effective against even insecticide-resistant disease-transmitting vectors, particularly targeting immature stages at low dosages of 1-30 ppm (Benelli, 2016). The longevity of essential oil strength is increased by the production of plant-derived nanoparticles and non-encapsulation chemicals by the process of slow release for long-term protection against mosquito bites (Maia and Moore, 2011). Phytochemicals might also be utilized to control mosquito vectors by changing their behaviour and perhaps replacing pyrethroids in bed net impregnation (Deletre *et al.*, 2015). However, the limitation of utilization of phytochemicals in IMM is poor characterization and inefficiency in

Table 1: Efficacy of botanical extracts among various solvents in term of LC₅₀ against the major vector Mosquito's larvae

S. No.	Reference	Target Mosquitoes	Plants Scientific Name (family)	Plant Parts Used	LC ₅₀ (PPM)				
					Methanol	Ethanol	Chloroform	Ethyl acetate	Petroleum Ether
1	Dakum <i>et al.</i> (2021)	<i>Anopheles stephensi</i>	<i>Hyptis suaveolens</i> (Lamiaceae)	Leaf	316.22	-	-	-	-
2	Sogan <i>et al.</i> (2020)	<i>Anopheles culicifacies</i>	<i>Spergula arvensis</i> (Caryophyllaceae)	Leaf	6.843	-	-	-	-
3	Shinde and Goyal (2020)	<i>Aedes aegypti</i>	<i>Datura inoxia</i> (Solanaceae)	Seed	72.44	-	-	-	-
4	Paul <i>et al.</i> (2020)	<i>Aedes aegypti</i>	<i>Andrographis paniculata</i> (Acanthaceae)	Stem, Leaves, Roots	154.94	197	-	-	-
			<i>Tinospora cordifolia</i> (Menispermaceae)	Stem, Leaves	181.63	162.88	-	-	-
5	Egunjobi and Okoye (2020)	<i>Anopheles gambiae</i>	<i>Datura erecta</i> (Solanaceae)	Leaf	-	76.94	-	-	-
			<i>Tridax procumbens</i> (Asteraceae)		-	214.22	-	-	-
			<i>Pennisetum purpureum</i> (Poaceae)		-	213.41	-	-	-
6	Al-Mekhlafi <i>et al.</i> (2020)	<i>Culex pipiens</i>	<i>Matricaria chamomilla</i> (Asteraceae)	Flower	-	-	-	287.1	-
7	Ali and Venkatesolu (2020)	<i>Anopheles stephensi</i>	<i>Saussurea costus</i> (Asteraceae)	Leaf	18.96	-	57.37	73.35	17.72
		<i>Aedes aegypti</i>			26.52	-	75.84	113.49	23.49
		<i>Culex quinquefasciatus</i>			35.85	-	84.14	136.34	50.12
		<i>Anopheles stephensi</i>		Root	7.96	-	29.84	39.45	14.26
		<i>Aedes aegypti</i>			10.79	-	49.56	86.95	22.42

		<i>Culex quinquefasciatus</i>			15.31		66.54	90.68	31.58
8	Raj and Shettu (2020)	<i>Aedes aegypti</i>	<i>Cymbopogon citratus</i> (Poaceae)	Leaf	Aqueous				Hexane
					299.98	164.31	256.59	244.24	196.37
			<i>Syzygium aromaticum</i> (Myrtaceae)		205.71	194.74	213.42	213.89	205.44
			<i>Eucalyptus deglupta</i> (Myrtaceae)		269.13	72.19	219.0	113.87	199.0
			<i>Azadirachta indica</i> (Meliaceae)		663.06	130.37	165.05	185.30	400.63
			<i>Tagetes erecta</i> (Asteraceae)		160.29	141.44	167.16	288.37	135.62
		<i>Culex quinquefasciatus</i>	<i>Cymbopogon citratus</i> (Poaceae)		270.37	140.27	233.08	220.99	184.46
			<i>Syzygium aromaticum</i> (Myrtaceae)		184.14	177.94	188.11	186.05	200.65
			<i>Eucalyptus deglupta</i> (Myrtaceae)		119.77	63.54	213.73	109.07	186.76
			<i>Azadirachta indica</i> (Meliaceae)		424.56	118.24	143.61	151.37	370.9
			<i>Tagetes erecta</i> (Asteraceae)		138.40	119.54	137.21	234.85	117.14
9	Shahat et al. (2020)	<i>Culex pipiens</i>	<i>Otostegia fruticosa</i> (Lamiaceae)	Leaf	683.28	-	261.13 (chlorobenzene)	585.48	129.92
10	Karthi et al. (2020)	<i>Aedes aegypti</i>	<i>Rhizophora mucronata</i> (Rhizophoraceae)	Leaf			Acetone		Petro-benz
					154	-	113	305	502
		<i>Anopheles stephensi</i>			415	-	378	427	504
		<i>Culex quinquefasciatus</i>			295	-	129	378	584

defining the structure of active toxic components as major reasons for the failure of bioactive toxic phytochemicals to migrate from the laboratory to the field.

The following stages might be recommended for any study design using phytochemicals to follow a futuristic strategy for the long-term objective of producing a green biopesticide: (i) screening of floral biodiversity for crude plant extracts with larvicidal efficacy against mosquitoes, (ii) preparation of plant solvent extracts ranging from non-polar to polar compounds, as well as the identification of the most efficient solvent extract with their lethal dose, (iii) phytochemical examination of the solid residue and structure determination using appropriate techniques such as column and thin layer chromatography, nuclear magnetic resonance (NMR), and gas chromatography and mass spectrometry (GC-MS), (iv) study of the effect of the active substances on non-target organisms, and finally (v) to check the active principle in the field before being recommended for use in a vector control programme and commercial manufacture.

Conclusion

The rapid increase in the human population, the increasing rate of global warming, synthetic pesticide resistance in mosquitoes and lack of awareness among people, together with environmental change and the adoptability of vector mosquitoes are responsible for the results of an increase in the mosquito's population and mosquito-borne diseases. Excessive indiscriminate and continuous use of synthetic pesticides has resulted in the progress of resistance species of mosquito and adverse effects on the environment and human health. Synthetic insecticides also negatively influence the soil, water, air, and food, contaminating the food we eat and the environment we live in. To prevent all of the negative consequences, we must develop more effective techniques to eliminate these vectors from our ecosystem.

Traditionally, plant based products have been

used in human communities for managing insect pests. Several secondary metabolites present in plants serve as a defense mechanism against insect attacks. A wide range of plant species from various geographical locations have been shown to contain phytochemicals capable of inducing a variety of acute and chronic harmful effects on mosquitoes. They have also been proven to cause a major deleterious impact on mosquito fertility and hatching, as well as larvicidal effects.

Several difficulties to be explored and addressed include residual effectiveness, preservation of non-target species, and uniformity of testing techniques. The recent studies included in this review suggest that plant botanicals and their components are extensively used as culinary herbs and spices, as well as pharmaceuticals. It will help to prevent the disposal of thousands of tonnes of pesticides into the environment and provide a safe environment to live in. Combined action of phytochemicals might be more promising for reducing the negative impacts and the development of mosquito resistant species.

Acknowledgements

The authors are thankful to Head of Department of Zoology, DDU Gorakhpur University, Gorakhpur (U.P.) for providing the facilities for this study.

References

- Ali SI and Venkatesalu V. (2020) Evaluation of the larvicidal potential of root and leaf extracts of *Saussurea costus* (Falc.) Lipsch. against three mosquito vectors: *Anopheles stephensi*, *Aedes aegypti*, and *Culex quinquefasciatus*. Rev Soc Bras Med Trop. 53: e20190018.
- Al-Mekhlafi FA, Abutaha N, Al-Malki AM and Al-Wadaan M. (2020) Inhibition of the growth and development of mosquito larvae of *Culex pipiens* L. (Diptera: Culicidae) treated with extract from flower of *Matricaria chamomilla* (Asteraceae). Entomol Res. 50(3): 138-145.
- Ault SK. (1994) Environmental management: A re-emerging vector control strategy. Am J Trop Med Hyg. 50 (Suppl): 35-49.
- Awad, OM and Shimaila A. (2003) Operational use of neem oil as an alternative anopheline larvicide. Part A: Laboratory and field efficacy. Eastern

- Mediterranean Hlth J. 9(4): 637.
- Barnard DR. (1999) Repellency of essential oils to mosquitoes (Diptera: Culicidae). J Med Entomol. 36: 625-629.
- Bayer Environmental Science (2003) Mosquito the vector for malarial and Filaria. www.ibl.gov.ERSP/generalinfo/contact.html.
- Benelli G. (2015) Plant-borne ovicides in the fight against mosquito vectors of medical and veterinary importance: A systematic review. Parasitol Res. 114: 3201-3212.
- Benelli G, Rajeswary M and Govindarajan M (2018) Towards green oviposition deterrents? Effectiveness of *Syzygium lanceolatum* (Myrtaceae) essential oil against six mosquito vectors and impact on four aquatic biological control agents. Environ Sci Pollut Res Int. 25(11): 10218-10227.
- Brogdon WG and Barber AM. (1990) Fenitrothion-deltamethrin cross-resistance conferred by esterases in Guatemalan *Anopheles albimanus*. Pest Biochem Physiol. 37: 130-139.
- Brown AWA. (1986) Insecticide resistance in mosquitoes: a pragmatic review. J Am Mosq Control Assoc. 2:123-140.
- Campbell FL Sullivan WW and Smith LN. (1993) The relative toxicity of nicotine, anabasine, methyl anabasine and lupinine for culicine mosquito larvae. J Econ Entomol. 26: 500.
- Cavalcanti ESB, Morais SM, Lima MAA and Santana EWP. (2004) Larvicidal activity of essential oils from Brazilian plants against *Aedes aegypti* L. Mem Inst Oswaldo Cruz. 99: 541-544.
- Chaubey MK. (2019) Essential oils as green pesticides of stored grain insects, Eur J Biol Res. 9(4): 202-244.
- Chavasse DC and Yap HH. (1997) Chemical methods for the control of vectors and pests of public health importance. WHO/CTD/WHOPES/97.2.WHO, Geneva.
- Chowdhury N, Ghosh A and Chandra G. (2008) Mosquito larvicidal activities of *Solanum villosum* berry extract against the dengue vector *Stegomyia aegypti*. BMC Complement Altern Med. 8: 10.
- Da Silva GNS, Trindade FTT, dos Santos F, Gosmann G, Silva AA and Gnoatto SCB. (2016) Larvicidal activity of natural and modified triterpenoids against *Aedes aegypti* (Diptera: Culicidae). Pest Manag Sci. 72(10): 1883-1887.
- Dakum YD, Amajoh CN, Ombugadu A, Istifanus G, Agwom F, Joseph ST, Jwanse IR, Lapang PM, Kpodorah SW and Pam DD. (2020) Larvicidal efficacy and GC-MS analysis of *Hyptis suaveolens* leaf extracts against *Anopheles*. Int J Biochem Res Rev. 30(1): 8-19.
- Das PK and Amalraj DD. (1997) Biological control of malaria vectors. Indian J Med Res. 106: 174-197.
- David JP, Rey D, Pautou MP and Meyran JC. (2000) Differential toxicity of leaf litter to dipteran larvae of mosquito developmental sites. J Invertebr Pathol. 75: 9-18.
- Dawet A, Ikani AG and Yakubu DP. (2016) Larvicidal effect of *Hyptis suaveolens* and *Chenopodium ambrosioides* on *Anopheles* mosquito larvae. Int J Sci Engineer Res. 7(1): 456-470.
- Deletre E, Chandre F, Barkman B, Menut C and Martin T. (2016) Naturally occurring bioactive compounds from four repellent essential oils against *Bemisia tabaci* whiteflies. Pest Manag. Sci. 72: 179-189.
- Egunjobi FB and Okoye IC. (2020) Ovicidal and larvicidal activities of ethanolic leaf extracts of three botanicals against the malaria vector - *Anopheles gambiae*. Int Ann Sci. 9(1): 111-121.
- Elimam AM, Elmalik KH and Ali FS. (2009) Efficacy of leaves extract of *Calotropis procera* Ait. (Asclepiadaceae) in controlling *Anopheles arabiensis* and *Culex quinquefasciatus* mosquitoes. Saudi J Biol Sci. 16: 95-100.
- Enan E. (2001) Insecticidal activity of essential oils: octopaminergic sites of action. Comp Biochem Physiol. C130: 325-337.
- Eshgy N. (1977) Tolerance of *Anopheles stephensi* to malathion in the province of Fars, Southern Iran. Mosq News 38: 580-583.
- Fournier D and Mutero A. (1994) Modification of acetylcholinesterase as a mechanism of resistance to insecticides. Comp Biochem Physiol C Pharmacol Toxicol Endocrinol. 108(1): 19-31.
- Ghosh A, Chowdhury N and Chandra G. (2012) Plant extract as potential mosquito larvicides. Indian J Med Res. 135(23): 581-598.
- Govindarajan M, Mathivanan T, Elumalai K, Krishnappa K and Anandan A. (2011) Mosquito larvicidal, ovicidal, and repellent properties of botanical extracts against *Anopheles stephensi*, *Aedes aegypti*, and *Culex quinquefasciatus* (Diptera: Culicidae). Parasitol Res. 109: 353-367.
- Govindarajan M, Rajeswary M, Hoti SL, Bhattacharyya A and Benelli G. (2016a) Eugenol, α -pinene and β -caryophyllene from *Plectranthus barbatus* essential oil as eco-friendly larvicides against malaria, dengue and Japanese encephalitis mosquito vectors. Parasitol Res. 115(2): 807-815.
- Govindarajan M, Rajeswary M and Benelli G. (2016b) Chemical composition, toxicity and non-target effects of *Pinus kesiya* essential oil: an eco-friendly and

- novel larvicide against malaria, dengue and lymphatic filariasis mosquito vectors. *Ecotoxicol Environ Saf.* 129: 85-90.
- Govindarajan M, Rajeswary M, Hoti SL and Benelli G. (2016c) Larvicidal potential of carvacrol and terpinen-4-ol from the essential oil of *Origanum vulgare* (Lamiaceae) against *Anopheles stephensi*, *Anopheles subpictus*, *Culex quinquefasciatus* and *Culex tritaeniorhynchus* (Diptera: Culicidae). *Res Vet Sci.* 104: 77-82.
- Govindarajan M, Rajeswary M, Arivoli S, Tennyson S and Benelli G. (2016d) Larvicidal and repellent potential of *Zingiber nimmonii* (J. Graham) Dalzell (Zingiberaceae) essential oil: an eco-friendly tool against malaria, dengue, and lymphatic filariasis mosquito vectors? *Parasitol Res.* 115(5):1807-1816.
- Govindarajan M, Rajeswary M and Benelli G (2016e) δ -Cadinene, calarene and δ -4-carene from *Kadsura heteroclita* essential oil as novel larvicides against malaria, dengue and filariasis mosquitoes. *Comb Chem High Throughput Screen.* 19(7): 565–571.
- Gubler DJ. (1998) Resurgent vector-borne diseases as a global health problem. *Emerg Infect Dis.* 4: 1-9.
- Hardin JA and Jackson FLC. (2009) Applications of natural products in the control of mosquito-transmitted diseases. *Afr J Biotechnol.* 8: 7373-7378.
- Hemalatha P, Elumalai D, Janaki A, Babu M, Velu K, Velayutham K and Kaleena PK. (2015). Larvicidal activity of *Lantana camara aculeata* against three important mosquito species. *J Entomol Zool Stud.* 3(1):174-181.
- Hemant GM and Shinde LV. (2020) Mosquito larvicidal efficacy of methanolic extract from seeds of *Datura innoxia* Mill. against *Aedes aegypti* (Linn.) with insight into GC-MS analysis. *J Entomol Res.* 44: 107-112.
- Hemingway J. (1983) Biochemical studies on malathion resistance in *Anopheles arabiensis* from Sudan. *Trans R Soc Trop Med Hyg.* 77: 477-480.
- Hemingway J and Georgiou GP. (1983) Studies on the acetylcholinesterase of *Anopheles albimanus* resistant and susceptible to organophosphate and carbamate insecticides. *Pesticide Biochem Physiol.* 19: 167-171.
- Hemingway J, Malcolm CA, Kissoon KE, Boddington RG, Curtis CF and Hill N. (1985) The biochemistry of insecticide resistance in *Anopheles sacharovi*: comparative studies with a range of insecticide susceptible and resistant *Anopheles* and *Culex* species. *Pesticide Biochem Physiol.* 24: 68-76.
- Hemingway J, Small GJ, Monro A, Sawyer BV and Kasap H. (1992) Insecticide resistance gene frequencies in *Anopheles sacharovi* populations of the Cukurova plain, Adana province, Turkey. *Med Vet Entomol.* 6: 342-348.
- Herath PRJ, Hemingway J, Weerasinghe IS and Jayawardena KGI. (1987) The detection and characterization of malathion resistance in field populations of *Anopheles culicifacies* B in Sri Lanka. *Pesticide Biochem Physiol.* 29: 157-162.
- Ignacimuthu S. (2000) The root of botanicals in combating mosquitoes. *Proc Symp Recent Trends Combating Mosquitoes*, Loyola College, Chennai, India.
- Isman MB. (1997) Neem and other botanical insecticides: barriers to commercialization. *Phytoparasitica* 25: 339-344.
- Isman MB. (2015) A renaissance for botanical insecticides? *Pest Manag Sci.* 71(12):1587-1590.
- Jacobson M and Crosby DG. (1971) Naturally occurring insecticides. Marcel Dekker Inc. New York, p. 585.
- James AA. (1992) Mosquito molecular genetics: the hands that feed bite back. *Science* 257: 37-38.
- Joseph CC, Ndoile MM, Malima RC and Nkunya MH. (2008) Larvicidal and mosquitocidal extracts, a coumarin, isoflavonoids and pterocarpan from *Neorautanenia mitis*. *Trans R Soc Trop Med Hyg.* 98(8): 451-455.
- Kalita B, Bora S and Sharma AK. (2013) Plant essential oils as mosquito repellent –A review. *Int J Res Develop Pharm Life Sci.* 3(1): 741-747.
- Karthi S, Uthirarajan K, Manohar V, Venkatesan M, Chinnaperumal K, Srinivasan PV and Krutmuan P. (2020) Larvicidal enzyme inhibition and repellent activity of red mangrove *Rhizophora mucronata* (Lam.) leaf extracts and their biomolecules against three medically challenging arthropod vectors. *Molecules* 25(17): 3844.
- Kawada H, Higa Y, Nguyen YT, Tran SH, Nguyen HT and Takagi M. (2009) Nationwide investigation of the pyrethroid susceptibility of mosquito larvae collected from used tires in Vietnam. *PLoS Negl Trop Dis.* 3(3): e391.
- Khanna VG and Kannabiran K. (2007) Larvicidal effect of *Hemidesmus indicus*, *Gymnema sylvestre*, and *Eclipta prostrata* against *Culex quinquefasciatus* mosquito larvae. *Afr J Biotechnol.* 3: 307-311.
- Kishore N, Mishra BB, Tiwari VK and Tripathi V. (2011) A review on natural products with mosquitocidal potentials. In: *Opportunity, challenge and scope of natural products in medicinal chemistry*, (ed.) Tiwari VK., Kerala Trans World Publishers, Research Signpost, pp. 223–253.
- Koureas M, Tsakalof A, Tsatsakis A and Hadjichristodoulou C. (2012) Systematic review of biomonitoring studies to determine the association between exposure to organophosphorus and

- pyrethroid insecticides and human health outcomes. *Toxicol Lett.* 210: 155-168.
- Kumar G, Karthik L, Rao BVK, Kirthi VA, Jayaseelan C and Rehuman AA. (2012) Phytochemical composition, mosquito larvicidal, ovicidal and repellent activity of *Calotropis procera* (Ait.) R. Br. against *Culex tritaeniorhynchus* and *Culex gelidus*. *Bangladesh J Pharmacol* 7(1): 63-69.
- Kumar S, Kumar S and Kapinder (2021) Evolution of larvicidal potential of indigenous plants of eastern Uttar Pradesh. Present status and future trends in entomological and wildlife studies. PENWI -21, Himanshu Publications, pp. 43-48.
- Lalthazuali and Nisha M. (2017) Mosquito repellent activity of volatile oils from selected aromatic plants. *Parasitol Res.* 116(2): 821-825.
- Lawler SP and Lanzaro GC. (2005) Managing Mosquitoes on the Farm, UC ANR, Publication: 8158.
- Lee BH, Lee SE, Annis PC, Pratt SJ, Park BS and Tumaalii F. (2002) Fumigant toxicity of essential oils and monoterpenes against the red flour beetle, *Tribolium castaneum* Herbst. *J Asia-Pacific Entomol.* 5(2): 237-240.
- Lixi S, Dong H, Guo C, Qian J, Sun J, Ma L and Zhu C. (2006) Larvicidal activity of extracts of *Ginkgo biloba* exocarp for three different strains of *Culex pipiens pallens*. *J Med Entomol.* 43(2): 258-261.
- Macedo ME, Consoli RA, Grandi TS, dos Anjos AM, de Oliveira AB, Mendes NM, Queiróz RO and Zani CL. (1997) Screening of Asteracea plant extract for larvicidal activity against *Aedes fluviatilis*, *Mem Inst Oswaldo Cruz.* 92: 565-570.
- Maia MF and Moore SJ. (2011) Plant based insect repellents: a review of their efficacy, development and testing. *Malaria J.* 10: 511.
- Malik RB, Malik KM, Balakrishnan N and Suresh B. (2013) Evaluation of larvicidal activity of the different extracts against important species of mosquito: *Anopheles stephensi*. *J Parasitol Vector Biol.* 6(1): 11-15.
- Melrose J, Perroy R and Careas S. (2015) World population prospects. United Nation. 1: 578-592.
- Nerio LS, Olivero-Verbel J and Stashenko E. (2010) Repellent activity of essential oils: A review. *Bioresource Technol.* 101(1): 372-378.
- Nisha M, Anitha MG, Bala TSL, Sivakumar SM, Narmadha R and Kalyanasundaram M. (2009) Larvicidal activity of *Saraca indica*, *Nyctanthes arbortristis*, and *Clitoria ternatea* extracts against three mosquito vector species. *Parasitol Res.* 104(5): 1017-1025.
- Pal R. (1994) WHO/CMR program of genetic control of mosquito in India. In: The use of genetics in insect control, (eds.) Pal R. and Whitten M.J., Elsevier, North Holland.
- Paul A, Raj VS, Vibhuti A and Pandey RP. (2020) Larvicidal efficacy of *Andrographis paniculata* and *Tinospora cordifolia* against *Aedes aegypti*: A Dengue vector. *Pharmacog Res.* 12(4): 352-360.
- Pavela R. (2005) Insecticidal activity of some essential oils against larvae of *Spodoptera littoralis*. *Fitoterapia* 76: 691-696.
- Pavela R. (2015) Essential oils for the development of eco-friendly mosquito larvicides: A review. *Ind Crop Prod.* 76:174-187.
- Pavela R and Benelli G. (2016) Essential oils as ecofriendly biopesticides? Challenges and constraints. *Trends Plant Sci.* 21(12): 1000-1007.
- Pavela R, Maggi F, Lupidi G, Mbuntcha H, Woguem V, Womeni HM, Barboni L, Tapondjou LA and Benelli G. (2017) *Clausena anisata* and *Dysphania ambrosioides* essential oils: from ethno-medicine to modern uses as effective insecticides. *Environ Sci Poll Res.* 25: 10493-10503.
- Raghavendra K, Barik TK, Reddy BP, Sharma P and Dash AP. (2011) Malaria vector control: from past to future. *Parasitol Res.* 108 (4): 757-779.
- Raj SHC and Shettu N. (2020) Larvicidal activity of crude extracts and fractions of native plants against *Aedes aegypti* L. and *Culex quinquefasciatus* Say. (Diptera: Culicidae). *Int Res J Plant Sci.* 11(Spl. 1): 1-7.
- Rathy MC, Sajith U and Harilal CC. (2014) Plant diversity for mosquito control: A preliminary study. *Int J Mosquito Res.* 2(1): 29-33.
- Rattan RS. (2010) Mechanism of action of insecticidal secondary metabolites of plant origin. *Crop Protect.* 2: 913-920.
- Raveen R, Kamakshi KT, Deepa M, Arivoli S and Tennyson S. (2014) Larvicidal activity of *Nerium oleander* L. (Apocynaceae) flower extracts against *Culex quinquefasciatus* Say (Diptera: Culicidae). *Int J Mosquito Res.* 1(1): 38-42.
- Redwane A, Lazrek HB, Bouallam S, Markouk M, Amarouch H and Jana M. (2002) Larvicidal activity of extracts from *Quercus Lusitania* var *infectoria* galls (Oliv). *J Ethnopharmacol.* 79: 261-263.
- Reinert JF. (2000) New classification of composite genus *Aedes*. *J Am Mosquito Cont Assoc.* 16: 175-188.
- Rey D, Cuany A, Pautou M and Meyran J. (1999) Differential sensitivity of mosquito taxa to vegetable tannins. *J Chem Ecol.* 25: 537-548.
- Roy DN, Goswami R and Pal A. (2017) The insect repellents: A silent environmental chemical toxicant to the health. *Environ Toxicol Pharmacol.* 50:91-102.

- Sathishkumar M and Maneemegalai S. (2008) Evaluation of larvicidal effect of *Lantana camara* Linn against mosquito species *Aedes aegypti* and *Culex quinquefasciatus*. Adv Biol Res. 2(3-4): 39-43.
- Schofield CJ. (1993) The politics of malaria vector control. Bull Ent Res. 83: 1-4.
- Shaalan EAS, Canyon D, Younes MWF, Abdel-Wahab H and Mansour AH. (2005) A review of botanical phytochemicals with mosquitocidal potential. Environ Int. 31(8):1149-1166.
- Shahat MAM, Elsheikh T, Hammad K and Hasaballah A. (2020) Effect of some plant extracts on the biochemical parameters, AChE and GST activities of the mosquito, *Culex pipiens* L. (Diptera: Culicidae). Egypt Acad J Biol Sci E Med Entomol Parasitol. 12(2): 69-80.
- Shivakumar MS, Srinivasan R and Natarajan D. (2013) Larvicidal potential of some Indian medicinal plant extracts against *Aedes aegypti* (L.). Asian J Pharm Clin Res. 6(3): 77-80.
- Sogan N, Kala S, Kapoor N and Nagpal BN. (2020) Phytochemical analysis of *Spergula arvensis* and evaluation of its larvicidal activity against malarial vector *Anopheles culicifacies*. South Afr J Botany 137: 351-358.
- Srivastava VK, Rai M and Singh A. (2002) The effect of temperature variation on the synthetic pyrethroids susceptibility status of *Culex quinquefasciatus* in Gorakhpur district (U.P.). Ann Entomol. 20(1): 17-19.
- Times of India (2016) Encephalitis to be made "Notifiable disease": Government (published on 11 Aug 2016).
- Tripathi AK, Sha W, Shulaev V, Stins MF and Sullivan DJ (2009) *Plasmodium falciparum*-infected erythrocytes induce NF- κ B regulated inflammatory pathways in human cerebral endothelium. Blood 114(19): 4243-4252.
- Uttinger J, Tozan Y and Singer BH. (2001) Efficacy and cost-effectiveness of environmental management for malaria control. Trop Med Int Hlth. 6(9): 677-687.
- Vatandoost H, McCaffery AR and Townson H. (1996) An electrophysiological investigation of target site insensitivity mechanisms in permethrin-resistant and susceptible strains of *Anopheles stephensi*. Trans R Soc Trop Med Hyg. 90: 216.
- Wiseman Z and Chapagain BP. (2005) Larvicidal effects of aqueous extracts of *Balanites aegyptiaca* (desert date) against the larvae of *Culex pipiens* mosquitoes. Afr J Biotechnol. 4(11): 1351- 1354.
- World Health Organization (1992) Vector resistance to pesticides. Fifteenth report of the WHO expert committee on vector biology and control. Technical Report Series, 818: 1-62.
- World Health Organization (1996) Report of the WHO informal consultation on the evaluation on the testing of insecticides, CTD/WHO PES/IC/96.1. Geneva: p. 69.
- World Health Organization (2005) Guidelines for laboratory and field testing of mosquito larvicides, Communicable disease control, Prevention and eradication, WHO pesticide evaluation scheme. WHO, Geneva p. 69.
- World Health Organization (2021) World malaria report 2021.
- Zaim M and Guillet P. (2002) Alternative insecticides: an urgent need. Trends Parasitol. 18: 161-163.