

Influence of Oral Administration of a High-Calcium Solution into a Marine Teleost (Nibbler Fish) and a Freshwater Teleost (Goldfish) on Their Plasma Calcium Levels

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Abstract: In this study, the influence of the oral administration of a high-calcium solution on plasma calcium levels in a marine teleost has been investigated and compared with those in a freshwater teleost. The calcium consomme solution (calcium concentration was 0.6 M) was administered into each fish at a rate of 10 µl/g body weight. Blood samples were collected from the caudal vessels of each individual with heparinized syringes at 3, 9, and 24 h after injection. Thereafter, plasma calcium concentrations were measured. In nibbler fish (marine teleost), the plasma calcium levels tended to increase at 3 h after the injection of a high-calcium consomme solution, although there was no significant difference between experimental and control groups. At 9 and 24 h after injection, plasma calcium levels in nibbler fish did not change as compared with those in control fish. In goldfish (freshwater teleost), plasma calcium concentrations increased significantly at 3 and 9 h after the administration of a high-calcium consomme solution as compared with those of control fish. At 24 h after injection, plasma calcium concentrations in goldfish decreased to the control levels. To confirm the above results in nibbler fish, solutions of different calcium concentrations (0.1 and 1 M) were injected orally into nibbler fish. As a result, the ingested calcium (0.1 and 1 M) consomme solutions did not influence plasma calcium levels in nibbler fish as compared with those in control fish at 3 h after injection. It has been known that seawater-adapted teleosts produce insoluble calcium precipitates to excrete via the intestine extracalcium ingested by drinking seawater. The calcium precipitates were formed by a chemical reaction of Mg²⁺ and Ca²⁺ derived from ingested seawater with bicarbonate (HCO3-). In nibbler fish, Ca2+ ingested orally may bind to HCO3- and then form calcium precipitates. Therefore, we strongly suggest that the regulation of orally administered calcium is different between seawater- and freshwater-adapted teleosts.

Keywords: High-calcium solution, Plasma calcium, Calcium precipitates, Oral administration, Nibbler fish, Goldfish, Seawater adaptation

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Introduction

It is well-known that calcium is an essential mineral for maintaining cell viability and plays vital role in animals' life (Srivastav et al., 2000). In all vertebrates, blood calcium levels are strictly kept at a constant concentration (around 2.5 mM) in spite of changing the internal milieu or external environment (Dacke, 1979; Bentley, 1998). The diet is a physiologically important source of calcium, although teleosts have the ability to absorb a large part of the necessary calcium from the environmental water (Dacke, 1979; Bentley, 1998). Therefore, we previously examined the effects of the oral administration of a highcalcium solution (consomme soup containing CaCl₂) into goldfish (*Carassius auratus*) (Sasayama et al., 1996) and eels (Anguilla *japonica*) (Suzuki *et al.*, 1999) on their plasma calcium levels. As a result, plasma calcium concentrations increased significantly in both goldfish (Sasayama et al., 1996) and eels (Suzuki et al., 1999). In freshwater teleosts such as goldfish and eels, thus, calcium contained in the diet is an important source in a low calcium environment, such as fresh water.

However, the influence of the oral administration of a calcium solution into marine teleosts on plasma calcium levels has not yet been fully elucidated because marine teleosts inhabit seawater that contains around 3–5 times the calcium in their blood (Björnsson and Nilsson, 1985; Suzuki *et al.*, 1999; Sato *et al.*, 2017; Zanaty *et al.*, 2020). In the present study, we examined the effects of the oral administration of a high-calcium consomme solution on the plasma calcium levels of marine teleost (nibbler fish) and compared them with those of freshwater teleost (goldfish).

Materials and Methods

Animals

Nibbler fish (*Girella punctata*) (both sexes, n = 16, 263 \pm 22.2 g) (Fig. 1A) were captured by fishing in Tsukumo Bay of the Noto Peninsula (Ishikawa Prefecture). After acclimation for 2 weeks, these fish were used in the present experiments. Freshwater teleost, goldfish (*Carassius auratus*) (both sexes, n = 16, 21.0 \pm 1.44 g) (Fig. 1B) were purchased from a commercial source (Higashikawa Fish Farm, Yamatokoriyama, Japan) and used in the present study.

All experimental procedures were conducted in accordance with the Guide for the Care and Use of Laboratory Animals prepared by Kanazawa University.

Experiment I: Comparison of the oral administration of a high-calcium solution on the plasma calcium levels of marine and freshwater teleosts:

Nibbler fish (marine teleost) were divided into two groups. One group was anesthetized with 0.03% tricaine (Tokyo Chemical Industry Co., Ltd., Tokyo, Japan) and administered a highcalcium solution directly into the digestive tract through the oral cavity by an oral sonde with a syringe (Fig. 1C). The calcium solution consisted of a consomme cube and CaCl₂, and the final calcium concentration was 2.4 g Ca/100 ml (0.6 M). The high-calcium consomme solution was given to each fish at a rate of 10 μ l/g body weight. The other group (control) was given only the consomme solution in the same way. In the case of freshwater teleost (goldfish), fish were divided into two groups and administered a highcalcium consomme solution (experimental) or a consomme solution (control) into the

digestive tract through the oral cavity by an oral sonde with a syringe (Fig. 1C) under anesthesia.



Fig. 1. Experimental fish (A: nibbler fish; B: goldfish) and equipment for the injection of calcium consomme solution (C).

Blood samples were collected from the caudal vessels of each individual with heparinized syringes at 3, 9, and 24 h after injection from nibbler fish (control and experimental) and goldfish (control and experimental) under anesthesia. The blood was collected into a 1.5 mL tube. Thereafter, the blood was centrifuged at 15,000 rpm for 3 min. The separated plasma was immediately frozen and kept at -80 °C until use. The plasma total calcium levels (mg/dL) of fish were determined using an assay kit (Calcium E, FUJIFILM Wako Pure Chemical Corporation, Osaka, Japan) with a microplate reader (CORONA ELECTRIC Co., Ltd., Hitachinaka, Ibaraki, Japan) as described by Suzuki et al. (2011) and Sato et al. (2017).

Experiment II: Influence of the oral administration of different calcium solutions on plasma calcium levels in nibbler fish:

To confirm the above results in nibbler fish, consomme solutions of different calcium concentrations (0.1 and 1 M) were orally injected into nibbler fish. Nibbler fish were divided into three groups. Two groups were anesthetized with 0.03% of tricaine (Tokyo Chemical Industry Co., Ltd.) and administered

a 0.1M (first group) and 1M calcium (second group) consomme solution directly into the digestive tract through the oral cavity by an oral sonde with a syringe (Fig. 1C), as described in experiment I. The third group (control group) was given only the consomme solution in the same way.

Blood samples were collected from the caudal vessels of each individual with heparinized syringes at 3 h after injection under anesthesia. The blood was collected into a 1.5 mL tube. Thereafter, the blood was centrifuged at 15,000 rpm for 3 min. The separated plasma was immediately frozen and kept at -80 °C until use. The plasma total calcium levels (mg/dL) in nibbler fish were determined using an assay kit (Calcium E, FUJIFILM Wako Pure Chemical Corporation) with a microplate reader (CORONA ELECTRIC Co., Ltd.) as described by Suzuki *et al.* (2011) and Sato *et al.* (2017).

Statistical analysis

All results are expressed as the means \pm SE. The statistical significance between the control and experimental groups was assessed by student *t*-test. In all cases, the selected significance level was p< 0.05.

Results

Experiment I: Comparison of the oral administration of a high-calcium solution on the plasma calcium levels of marine and freshwater teleosts:

In nibbler fish (marine teleost), plasma calcium levels tended to increase at 3 h after the injection of a high-calcium consomme solution, although there was no significant difference between experimental and control groups (Fig. 2A). At 9 h (Fig. 3A) and 24 h (Fig. 4A) after injection, plasma calcium levels in nibbler fish did not change as compared with those in control fish.



Fig. 2. Comparison of the oral administration of a calcium (0.6 M) consomme solution on the plasma calcium levels of nibbler fish (A) and goldfish (B). At 3 hours after administration, plasma calcium concentrations of nibbler fish and goldfish were measured. Each n = 8. **: p < 0.01. CTRL: control group; Ca Adm.: calcium administration group.



Fig. 3. Comparison of the oral administration of a calcium (0.6 M) consomme solution on the plasma calcium levels of nibbler fish (A) and goldfish (B). At 9 hours after administration, plasma calcium concentrations of nibbler fish and goldfish were measured. Each n = 8. *: p < 0.05. CTRL: control group; Ca Adm.: calcium administration group.

In goldfish (freshwater teleost), plasma calcium levels increased significantly at 3 h (Figure 2B) and 9 h (Fig. 3B) after the administration of a high-calcium consomme solution as compared with control goldfish injected with only a consomme solution. At 24 h after injection, plasma calcium concentration in goldfish decreased to the control levels (Fig. 4B). There was no significant difference between experimental and control groups at 24 h after injection (Fig. 4B).



Fig. 4. Comparison of the oral administration of a calcium (0.6 M) consomme solution on the plasma calcium levels of nibbler fish (A) and goldfish (B). At 24 hours after administration, plasma calcium concentrations of nibbler fish and goldfish were measured. Each n = 8. There was no significant difference between control and experimental groups in these fish. CTRL: control group; Ca Adm.: calcium administration group.



Fig. 5. Effect of the oral administration of calcium (A: 0.1 M; B: 1 M) consomme solution on plasma calcium levels in nibbler fish. At 3 hours after administration, plasma calcium concentrations were measured in nibbler fish. Each n = 3. There was no significant difference between control and experimental groups in nibbler fish. CTRL: control group; Ca Adm.: calcium administration group.

Experiment II: Influence of the oral administration of different calcium solutions on plasma calcium levels in nibbler fish:

To confirm the results of experiment I in nibbler fish, consomme solutions of different calcium concentrations (0.1 and 1 M) were orally injected into nibbler fish. At the most effective time-course (3 h after injection; Fig. 2 B) in goldfish, the plasma calcium in the injected nibbler fish was measured. In the ingested calcium concentrations (0.1 and 1 M), plasma calcium levels in nibbler fish were not changed as compared with those of control nibbler fish (Fig. 5).

Discussion

In the present study, we examined the effects of the oral administration of a high-calcium consomme solution on plasma calcium levels in nibbler fish (marine teleost) and compared them with those in goldfish (freshwater teleost). We found that, in nibbler fish, plasma calcium levels did not change significantly after the injection of a high-calcium (0.6 M) consomme solution, although plasma calcium concentrations in goldfish increased significantly at 3 and 9 h after the administration of a high-calcium (0.6 M) consomme solution, as compared with those in control goldfish. To confirm the above results in nibbler fish, consomme solutions of different calcium concentrations (0.1 and 1 M) were orally injected into nibbler fish. At the most effective time course (3 h after injection) in goldfish, the plasma calcium in the injected nibbler fish was measured. The ingested calcium (0.1 and 1 M) consomme solutions did not influence plasma calcium levels in nibbler fish as compared with those in control nibbler fish. We have previously reported on the effects of the oral administration of a high-calcium consomme solution (0.25 M) into freshwater teleosts such as goldfish (Carassius auratus) (Sasayama et al., 1996) and eels (Anguilla japonica) (Suzuki et al., 1999) on plasma calcium levels. As a result, plasma calcium concentrations increased significantly in both goldfish (Sasayama et al., 1996) and eels (Suzuki et al., 1999), as shown by our results (Figs. 2B, 3B). These facts, together with our obtained data, strongly suggest that the regulation of orally administered calcium seems to be different between seawater- and freshwater-adapted teleosts.

It has been known that marine teleosts excrete insoluble white feces, which is often referred to as calcium precipitates (Hickman, 1968; Walsh et al., 1991; Mekuchi et al., 2010; Whittamore et al., 2010; Mekuchi et al., 2013). The evacuation of ingested excess calcium by calcium precipitates was closely associated with seawater adaptation (Mekuchi et al., 2010). Insoluble calcium precipitates were formed and excreted through the digestive tract in seawater-adapted eels (Anguilla japonica) but not in freshwater-adapted eels (Mekuchi et al., 2010). In addition, Mekuchi et al. (2013) reported that calcium precipitates in the intestine of eels (Anguilla japonica) were formed by a chemical reaction of Mg²⁺ and Ca²⁺ derived from ingested seawater with bicarbonate (HCO_3^{-}) produced by the pancreas. In the European flounder (*Platichthys flesus*), furthermore, increasing the intestinal luminal Ca²⁺ concentration (10, 40, and 90 mM) caused a large elevation in intestinal HCO₃⁻ production in a dose-dependent manner (Cooper et al., 2010). In nibbler fish, therefore, it is possible that orally ingested Ca²⁺ binds to HCO₃- and then forms insoluble calcium precipitates.

These facts, together with our obtained results, suggest that the regulation of orally administered calcium is different between seawater- and freshwater-adapted teleosts. Marine teleosts, which live in a high-calcium environment (around 40 mg/dL) (Björnsson and Nilsson, 1985; Bentley, 1998), may have a specific mechanism to suppress the extracalcium obtained by food ingestion. In the intestine and pancreas of eels (*Anguilla*) *japonica*), the mRNA expression of bicarbonate transporters, such as solute carrier family 26, significantly increased when eels transferred from freshwater to seawater (Mekuchi *et al.*, 2013). To elucidate the regulation of orally administered calcium between seawater- and freshwater-adapted teleosts, we are planning to examine the expression levels of bicarbonate transporters after the injection of a high-calcium consomme solution in both freshwater- and seawater-adapted eels.

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References

- Bentley PJ. (1998) Hormones and calcium metabolism.
 In: Comparative Vertebrate Endocrinology, Bentley PJ (ed) 3rd ed. Cambridge University Press, Cambridge, pp 269-301
- Björnsson BTh and Nilsson S. (1985) Renal and extrarenal excretion of calcium in the marine teleost, *Gadus morhua*. Am. J. Physiol. Regul. Integr. Comp. Physiol. 248: R18-R22.
- Cooper CA, Whittamore JM and Wilson RW. (2010) Ca²⁺driven intestinal HCO₃- secretion and CaCO₃ precipitation in the European flounder *in vivo*: Influences on acid-base regulation and blood gas transport. Am. J. Physiol. Regul. Integr. Comp. Physiol. 298: R870-R876.
- Dacke CG. (1979) Calcium regulation in sub-mammalian vertebrates. Academic Press, London.
- Hickman CP Jr. (1968) Ingestion, intestinal absorption, and elimination of seawater and salts in the southern flounder, *Paralichthys lethostigma*. Can. J. Zool. 46: 457-466.
- Mekuchi M, Hatta T and Kaneko T. (2010) Mg-calcite, a carbonate mineral, constitutes Ca precipitates produced as a byproduct of osmoregulation in the

intestine of seawater-acclimated Japanese eel *Anguilla japonica*. Fish. Sci. 76: 199–205.

- Mekuchi M, Watanabe S and Kaneko T. (2013) Bicarbonate secreted from the pancreas contributed to the formation of Ca precipitates in Japanese eel, *Anguilla japonica*. J. Exp. Zool. Part A 319: 53-62.
- Sasayama Y, Abe I, Suzuki N and Hayakawa T. (1996) Plasma calcium and calcitonin levels at food intake in eels and goldfish. Zool. Sci. 13: 731-735.
- Sato M, Yachiguchi K, Motohashi K, Yaguchi Y, Tabuchi Y, Kitani Y, Ikari T, Ogiso S, Sekiguchi T, Hai TN, Huong DTT, Hoang NV, Urata M, Mishima H, Hattori A and Suzuki N. (2017) Sodium fluoride influences calcium metabolism resulting from the suppression of osteoclasts in the scales of nibbler fish *Girella punctata*. Fish. Sci. 83: 543–550.
- Srivastav Ajai K, Das VK, Srivastav SK and Suzuki N. (2000) Amphibian calcium regulation: Physiological aspects. Zool. Ploniae 45: 9-36.
- Suzuki N, Suzuki D, Sasayama Y, Srivastav Ajai K, Kambegawa A and Asahina K. (1999) Plasma calcium and calcitonin levels in eels fed a high calcium solution or transferred to seawater. Gen. Comp. Endocrinol. 114: 324-329.
- Suzuki N, Danks JA, Maruyama Y, Ikegame M, Sasayama Y, Hattori A, Nakamura M, Tabata MJ, Yamamoto T, Furuya R, Saijoh K, Mishima H, Srivastav Ajai K, Furusawa Y, Kondo T, Tabuchi Y, Takasaki I, Chowdhury VS, Hayakawa K and Martin TJ. (2011) Parathyroid hormone 1 (1-34) acts on the scales and involves calcium metabolism in goldfish. Bone 48: 1186-1193.
- Walsh PJ, Blackwelder P, Gill KA, Danulat E and Mommsen TP. (1991) Carbonate deposits in marine fish intestines: A new source of biomineralization. Limnol. Oceanogr. 36: 1227-1232.
- Whittamore JM, Cooper CA and Wilson RW. (2010) HCO₃secretion and CaCO₃ precipitation play major roles in intestinal water absorption in marine teleost fish *in vivo*. Am. J. Physiol. Regul. Integr. Comp. Physiol. 298: R877-R886.
- Zanaty MI, Sawada N, Kitani Y, Nassar HF, Mahmoud HM, Hayakawa K, Sekiguchi T, Ogiso S, Tabuchi Y, Urata M, Matsubara H, Takeuchi Y, Hattori A, Srivastav Ajai K, Amornsakun T and Suzuki N. (2020) Influence of benz[a]anthracene on bone metabolism and on liver metabolism in nibbler fish, *Girella punctata*. Int. J. Environ. Res. Public Health 17: 1391.